

7-12-00

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07/11/00
JC877 U.S. PTO

PATENT

Docket No. FWLPAT013US

JC836 U.S. PTO
09/613355
07/11/00

Box Patent Application
Commissioner of Patents and Trademarks
Washington, D.C. 20231

NEW APPLICATION TRANSMITTAL
(STANDARD FORM)

Transmitted herewith for filing is the patent application of

Inventor(s): Binie V. Lipps, Frederick W. Lipps

For (title): Synthetic Peptide for Neurological Disorders

1. Type of Application

This new application is for an Original Application.

2. Papers Enclosed Which Are Required For Filing Date Under 37 CFR 1.53(b) (Regular) or 37 CFR 1.153 (Design) Application

 18 Pages of specification

 3 Pages of claims

 1 Page of Abstract

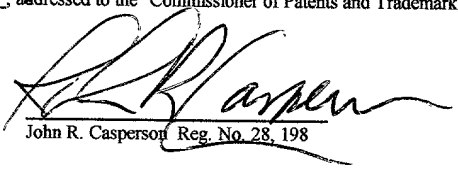
 0 Sheets of drawings

CERTIFICATION OF EXPRESS MAILING DATE

I hereby certify that this correspondence is being deposited by me with the United States Postal Service on 11 July 2000 in an envelope as "Express Mail, Post Office to Addressee",

bearing Label Number EK 7700965805, addressed to the "Commissioner of Patents and Trademarks, Washington, D.C. 20231".

Date 7-11-2000


John R. Casperson Reg. No. 28, 198

Send correspondence to:

John R. Casperson
PO Box 2174
Friendswood, Texas 77549

09/613355 07/11/00

3. Additional papers enclosed

☐ PTO 1449

☒ Preliminary Amendment

☒ Diskette with sequence listing

4. Declaration or oath

☒ Enclosed

executed by

☒ inventors.

5. Language

☒ English

6. Small Entity Statement(s)

☒ Verified Statement that this is a filing by a small entity attached.

7. Fee Payment Being Made At This Time

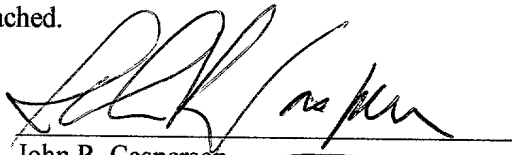
☒ Enclosed

<input checked="" type="checkbox"/> basic filing fee	\$ 345.00
4 independent claims over 3 @ \$39.00	\$ 156.00

Total fees enclosed	\$ 501.00
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8. Method of Payment of Fees

☒ A check in the amount of \$501.00 is attached.


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(281)-482-2961

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

22

Country	Year	Population (millions)	GDP (billion USD)	Urban population (%)	Life expectancy (years)	Infant mortality (per 1,000 live births)	Health expenditure (billion USD)	Health expenditure per capita (USD)
Algeria	2000	29.0	10.0	55.0	72.0	100.0	0.5	17.2
Algeria	2001	29.2	10.5	55.5	72.5	95.0	0.5	17.5
Algeria	2002	29.4	11.0	56.0	73.0	90.0	0.5	17.8
Algeria	2003	29.6	11.5	56.5	73.5	85.0	0.5	18.1
Algeria	2004	29.8	12.0	57.0	74.0	80.0	0.5	18.4
Algeria	2005	30.0	12.5	57.5	74.5	75.0	0.5	18.7
Algeria	2006	30.2	13.0	58.0	75.0	70.0	0.5	19.0
Algeria	2007	30.4	13.5	58.5	75.5	65.0	0.5	19.3
Algeria	2008	30.6	14.0	59.0	76.0	60.0	0.5	19.6
Algeria	2009	30.8	14.5	59.5	76.5	55.0	0.5	19.9
Algeria	2010	31.0	15.0	60.0	77.0	50.0	0.5	20.2
Algeria	2011	31.2	15.5	60.5	77.5	45.0	0.5	20.5
Algeria	2012	31.4	16.0	61.0	78.0	40.0	0.5	20.8
Algeria	2013	31.6	16.5	61.5	78.5	35.0	0.5	21.1
Algeria	2014	31.8	17.0	62.0	79.0	30.0	0.5	21.4
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Algeria	2016	32.2	18.0	63.0	80.0	20.0	0.5	22.0
Algeria	2017	32.4	18.5	63.5	80.5	15.0	0.5	22.3
Algeria	2018	32.6	19.0	64.0	81.0	10.0	0.5	22.6
Algeria	2019	32.8	19.5	64.5	81.5	5.0	0.5	22.9
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Algeria	2022	33.4	21.0	66.0	83.0	0.0	0.5	23.8
Algeria	2023	33.6	21.5	66.5	83.5	0.0	0.5	24.1
Algeria	2024	33.8	22.0	67.0	84.0	0.0	0.5	24.4
Algeria	2025	34.0	22.5	67.5	84.5	0.0	0.5	24.7
Algeria	2026	34.2	23.0	68.0	85.0	0.0	0.5	25.0
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Algeria	2038	36.6	29.0	74.0	91.0	0.0	0.5	28.6
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Algeria	2040	37.0	30.0	75.0	92.0	0.0	0.5	29.2
Algeria	2041	37.2	30.5	75.5	92.5	0.0	0.5	29.5
Algeria								

[] persons, concerns or organizations listed below

I acknowledge the duty to file, in this application or patent, notification of any change in status resulting in loss of entitlement to small entity status prior to paying, or at the time of paying, the earliest of the issue fee or any maintenance fee due after the date on which status as a small entity is no longer appropriate. (37 CFR 1.28(b)).

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application, any patent issuing thereon, or any patent to which this verified statement is directed.

Binie V. Lipps 7/10/00
Binie V. Lipps (date)

Frederick W. Lipps 7/10/00
Frederick W. Lipps (date)

PATENT

IN THE UNITED STATES PATENT AND TRADEMARK OFFICE

In re application of:
Binie V. Lipps
Frederick W. Lipps

Serial No.:

Filed:

For:
SYNTHETIC PEPTIDE FOR
NEUROLOGICAL DISORDERS

§ ATTY DCKT NO: FWLPAT013US
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§ Art Unit:
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§ Examiner:
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Commissioner of Patent and Trademarks
Washington, D.C. 20231

PRELIMINARY AMENDMENT

Sir:

Prior to calculating the filing fee for the captioned patent application, and solely for the purpose of reducing the filing fee, kindly enter the following amendment.

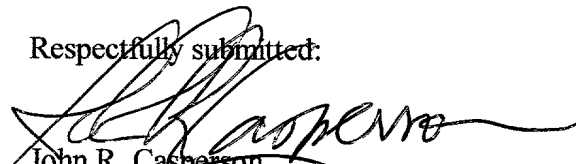
IN THE CLAIMS

Kindly cancel claims 2-5 and 8-10.

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Respectfully submitted:


John R. Casperson
Reg. No. 28,198

[illegible]

- (B) TYPE: AMINO ACID
(C) STRANDEDNESS: SINGLE
(D) TOPOLOGY: LINEAR
(ii) MOLECULE TYPE: PEPTIDE IN SEQ ID NO: 3
(iii) HYPOTHETICAL: NO
(iv) ANTI-SENSE:
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE: SYNTHETIC

- (A) ORGANISM:
(B) STRAIN:
(C) INDIVIDUAL ISOLATE:
(D) DEVELOPMENTAL STAGE:
(E) HAPLOTYPE:
(F) TISSUE TYPE:
(G) CELL TYPE:
(H) CELL LINE:
(I) ORGANELLE:

- (vii) SEQUENCE DESCRIPTION: SEQ ID NO: 3:
N L G E H P V C D S

(5) INFORMATION FOR SEQ ID NO: 4:

- (i) SEQUENCE CHARACTERISTICS:
(A) LENGTH: 5
(B) TYPE: AMINO ACID
(C) STRANDEDNESS: SINGLE
(D) TOPOLOGY: LINEAR
(ii) MOLECULE TYPE: PEPTIDE IN SEQ ID NO: 4
(iii) HYPOTHETICAL: NO
(iv) ANTI-SENSE:
(v) FRAGMENT TYPE:
(vi) ORIGINAL SOURCE: SYNTHETIC

- (A) ORGANISM:
(B) STRAIN:
(C) INDIVIDUAL ISOLATE:
(D) DEVELOPMENTAL STAGE:
(E) HAPLOTYPE:
(F) TISSUE TYPE:
(G) CELL TYPE:
(H) CELL LINE:
(I) ORGANELLE:

- (vii) SEQUENCE DESCRIPTION: SEQ ID NO: 4:
N L G E H

APPLICATION FOR PATENT

INVENTORS: BINIE V. LIPPS AND FREDERICK W. LIPPS

TITLE: SYNTHETIC PEPTIDE FOR NEUROLOGICAL DISORDERS

SPECIFICATION

BACKGROUND OF THE INVENTION

Field of the Invention

The invention relates to a synthetic peptide which mimics the biological properties of nerve growth factor (NGF) and is useful for treating neurological disorders.

Background of the invention

Nerve growth factor (NGF) was discovered more than forty years ago by Levi-Montalcini et al. in malignant tissues. Subsequently Cohen isolated NGF from snake venom and then from sub maxillary gland of mouse, a homolog to the snake venom gland. Since the discovery it was contemplated that NGF will be clinically useful to treat neurological disorders, like Alzheimer's (AD), Parkinson's (PD) and other neurological diseases. Over the years, several neurotrophic factors such as NT-3, bFGF, platelet derived growth factor, etc. were discovered. These factors regulate nerve cell growth and survival.

Research in animals has demonstrated that neurotrophic factors delivered to central nervous system can prevent or reverse neurodegeneration. Generally, neurotrophic factors cannot cross the blood-brain barrier due to their size and therefore, will not reach the brain when administered either orally or through injection. As a result, researchers have administered these proteins directly into the brain to determine their effectiveness in combating

neurodegenerative diseases. In animals, various neurotrophic factors administered through a hole drilled into the skull have been successful in restoring memory and stimulating nerve regeneration. In humans, nerve growth factor administered in similar way has improved memory in Alzheimer's disease patients.

5 A more convenient delivery system is required, to produce the beneficial effects that have been established for neurotrophic factors. The selective breakdown of the blood-brain barrier has not proven to be effective as yet. The most practical approach is to mimic the effects of neurotrophic factors by administration of an orally active compound that passes the blood-brain barrier and produces the effects of neurotrophic factors in the brain or turns on
10 the genes to produce neurotrophic factors at the site where they are needed in the brain.

AIT-082 is the first compound that has entered human clinical trials, which has been demonstrated to activate multiple genes in animals to produce three different neurotrophic factors (NGF, NT-3 and bFGF) in the specific areas of the brain associated with memory loss. In addition, AIT-082 has the advantage of being rapidly absorbed and active after oral
15 administration. However, its efficacy remains to be determined.

Appel proposed that selective neuronal degeneration may be caused by failure of target tissues to supply the necessary neurotrophic factor. A specific link between NGF and AD was first suggested by Hefti. It has been reported that NGF level rises in pathological situation such as hypoxic injury in adult rats. Data are already emerging to suggest an age-related reduction
20 in both NGF and its receptor in rat brain.

Naturally occurring bioactive peptides have been proposed for neurological disorders. Appel and Tomozawa 1991 isolated, extracted and purified three different neurotrophic factors from caudate putmen tissue of normal mammal, to treat amyotrophic lateral sclerosis (ALS), PD and AD. Heinrich produced recombinant human (h-NGF) made in Chinese hamster ovary

cells. (CHO). Lewis et al. 1992 proposed the use of insulin like growth factor for treating disorders enhancing the survival of non-mitotic cells.

Despite the previous failures to obtain NGF in animal sera, at Ophidia Products we have successfully isolated NGF from human and other animal sera, showing neurotrophic activity when tested on PC12 cells. In addition, we have isolated NGF from human saliva and urine. Furthermore, we have isolated NGF from the established cultures of eukaryotic cells; Chang cells (human liver), Vero (monkey kidney), pheochromocytoma PC12 (rat adrenal gland), neuroblastoma (human brain) and mouse myeloma (SP/2) cells.

According to Wells, 1996 the commonly held view that small synthetic peptides cannot mimic effects of large polypeptide ligands is by now considerably out of date. Several investigators have made synthetic NGF peptide derivatives which prevent neuronal death and show neurite outgrowth, the characteristic of the neurotrophic factor on PC12 cells. Longo et al. (1997) made cyclized peptides corresponding to beta loop region of NGF and found the highest activity corresponding a loop region 29-35 which is capable to interact with p75 receptor. According to them, to this date, no small molecule NGF agonist or partial agonists agents known to promote neurotrophic effects by acting via NGF receptors have been described.

A small molecule which behaves like NGF would be very desirable for treating neurological disorders, since it would overcome the blood-brain barrier. It would be capable of reaching the brain by most any route, for example, intramuscular, intravenous, buccal cavity or nasal insufflation could be used. It may avoid triggering antibody production.

A small molecule which behaves like NGF and can be synthetically produced would be even more desirable, since its production would be straightforward and inexpensive.

Objects of the Invention

The object of the invention is to provide a peptide consisting of in the range of 5 to 25 amino acids which mimics the activity of NGF. Such a peptide can be synthetically made in abundance to provide therapeutics for neurological disorders. Such a peptide would have a low molecular weight to enable it to reach the brain when administered by most any route.

SUMMARY OF THE INVENTION

The invention relates to a synthetic peptide consisting of at least the first five amino acids from the N-terminal of the sequence N L G E H P V C D S T D T W V. The synthetic peptide mimics the biological properties of nerve growth factor (NGF) consisting of 116 amino acids.

The peptide of the invention can be administered to a patient having a neurological disorder by various routes, including injection and orally. The peptide of the invention can reach the brain as a small molecule without blood-brain barrier problem, since it is small enough to cross the blood-brain barrier.

Antibodies made against the peptide of the invention have a higher binding affinity for NGF of human origin (termed H-NGF) than antibodies which were made against the 116 amino acid NGF derived from venom (V-NGF, the antibody being Anti-V-NGF). This fact evidences that the composition of the inventive peptide is a conserved domain of the activity of human NGF. Therefore, the inventive peptide is immunologically closer to H-NGF than V-NGF.

Antibodies made against the inventive peptide can be used assay NGF levels in human bodily fluids such as saliva and urine for diagnostic purposes without the necessity of extracting blood.

DETAILED DESCRIPTION OF THE INVENTION

The inventive peptides can be generally described as compositions of matter consisting of at least the first five amino acids from the N-terminal of the sequence N L G E H P V C D S T D T W V and no more than 25 amino acids total. Usually, the inventive peptides will contain no more than 20 amino acids, and preferably no more than 15 amino acids. I have named the inventive peptides ADESH.

ADESH which contains only a portion of the amino acid sequence is preferred. ADESH which contains only the first five amino acids of the sequence is termed AD-5. ADESH which contains only the first 10 amino acids of the sequence is termed AD-10, and is the preferred species. ADESH which contains only the first fifteen amino acids of the sequence is termed AD-15. AD-5, AD-10 and AD-15 have been tested on PC12 cells and found to be biologically active in producing neurite outgrowth.

ADESH constitutes a nerve growth factor preferably having in the range of 5 to 20 amino acids, and capable of crossing the blood-brain barrier. It can be effectively utilized by patients having a need of nerve growth factor by delivering it to the bloodstream. Examples of patients for whom ADESH treatment should be beneficial include victims of Alzheimer's disease (AD) and Parkinson's disease (PD). Suitable routes of administration include nasal insufflation, buccal administration, oral ingestion, and intramuscular injection. ADESH can also be injected directly into the blood stream.

ADESH mimics the biological properties of NGF derived from cobra venom. Venom-derived NGF is termed V-NGF. The property of V-NGF most interest which is mimicked by ADESH is the stimulation of neurite outgrowths.

The amino acid sequence of V-NGF derived from one species of cobra (*Naja naja*) venom is:

NH₂-

Glu-Asp-His-Pro-Val-His-Asn-Leu-Gly-Glu-His-Pro-Val-Cys-Asx-
Ser-Thr-Ash-Thr-Trp₂₀-Val-Gly-Val-Lys-Thr-Thr-Ala-Thr-Asn-Ile-
Lys-Gly-Ala-Ser-Val-Ser-Val-Met-Glu-Asn₄₀-Val-Asn-Leu-Asp-Asn-
Lys-Val-Tyr-Lys-Gln-Tyr-Phe-Phe-Glu-Thr-Lys-Cys-Arg-Asx-Ser₆₀-
Asx-Pro-Pro-Glx-Pro-Gly-Cys-Lys-Gly-Ile-Asx-Thr-Glx-His-Trp-
Asx-Ser-Tyr-Cys-Thr₈₀-Thr-Ser-Asn-Ser-Phe-Ile-Lys-Ala-Leu-Thr-
Met-Asx-Glx-Gly-Gln-Ser-Ala-Trp-Arg-Phe₁₀₀-Ile-Arg-Ile-Gix-Thr-
Ala-Cys-Val-Cys-Val-Ile-Thr-Lys-Lys-Gly-Asn-
COOH

In vivo, ADESH causes the immunized animal to produce an antibody which has a binding affinity to NGFs from human bodily fluids and human-origin eukaryotic cells which is higher than a binding affinity exhibited by an antibody produced in immunological response to V-NGF. Synthetic ADESH is equally active as the fragment of the native NGF. Antibodies made against ADESH (Anti-ADESH) react with V-NGF having 116 amino acids. However, antibodies made against V-NGF (Anti-V-NGF) react poorly with ADESH. This illustrates that the ten amino acids of ADESH are essentially important for the biological activity of neurite growth. Therefore, synthetic ADESH consisting of ten amino acids, especially, is a candidate for the treatment of neurological disorders instead of the entire NGF molecule.

In the past, there were failures to detect and/or quantify Human NGF (H-NGF) in human serum. Our research shows that H-NGF will can be quantified in vitro by contacting sample fluids with an Anti-ADESH. The contacting is preferably carried out so as to cause the antibody to react immunologically with the NGF contained in the sample fluid. The test has

been shown effective in quantifying H-NGF contained in samples of blood serum, saliva and urine.

EXPERIMENTAL AND RESULTS:

Purification of NGF from Snake venom:

Homogenous preparation of NGF was obtained by fractionating snake venom from *Naja kaouthia* by HPLC using ion exchange column and gradient Trizma-HCl buffer pH 7.4

Trypsin Digestion of Natural NGF:

Purified homogenous preparation of NGF was treated with trypsin dissolved in 0.1 M ammonium bicarbonate buffer pH 8.0. The NGF and the trypsin were mixed in 40:1 ratio, precisely 5 mg of NGF to 0.25 mg of trypsin. The mixture was incubated at 37° C to cause fragmentation at arginine and lysine sites. After 18 hours of incubation the reaction was stopped by cooling the mixture at 4° C.

Separation of Fragments from Trypsin Digest:

The trypsin digested fragments were separated on HPLC. The separation was done in two runs by loading half the mixture each time. Trypsin digested NGF resolved into ten different fragments. The fragments were collected individually and dialyzed against water using 500 daltons molecular weight cutoff tubing (Spectrum USA). The protein concentration of each fragment was measured by using Bio-Rad (USA) protein kit and the concentration of each fragment was adjusted to 100µg/ml with 0.05 M phosphate buffered saline (PBS).

Biological Activity of Fragments PC12 Cells:

The trypsin digested fragments in various concentrations were tested for neurite out growth on PC12 cells. Tissue culture plate having 24 wells were seeded with 10⁵ PC12 cells in serum free Dulbecco Modified Eagle's medium (DMEM). The results were read after 72 hours for neurite outgrowth. The fraction showing the most neurite outgrowth at the lowest concentration was sequenced for its amino acids composition. Sequencing was contracted out to the Protein Core

Laboratory of Baylor College of Medicine, Houston, Texas. The sequence for the fraction from the N-terminal was found to be: N L G E H P V C D S T D T W V.

Synthesis of ADESH:

Synthetic ADESH (AD-10) was constructed using the above amino acids sequence from N-terminal for ten amino acids N L G E H P V C D S. Two more versions of synthetic ADESH, termed AD-15 and AD-5, consisting of 15 and 5 amino acids respectively, were constructed; The peptides had the sequence: N L G E H P V C D S T D T W V for AD-15 and N L G E H for AD-5.

Production of Polyclonal Antibodies to ADESH in Mice:

There is a perception that small synthetic peptides do not generate antibodies on injection into animals. However, synthetic peptide can generate antibodies if it is tagged with a complete protein, before injecting to the animal. Landsteiner coined the term hapten for a low molecular weight, chemically defined compound which could induce antibody formation only when coupled to larger carrier protein molecule before injecting. Thereby, the injected animal makes antibodies to both the hapten and the carrier protein.

Synthetic chemically defined ADESH comprising fifteen, ten or five amino acids can be considered as haptens and therefore, theoretically should not induce antibodies if injected without a carrier protein. However, for our other projects, we have succeeded generating antibodies in mice for a synthetic peptide consisting of five amino acids.

Adult Balb/C mice were used for immunization. The mice were used in compliance with the US Public Health Service Policy on humane care and use of animals. First injection consisted of the mixture 100 μ g of each version of ADESH in 0.1 ml mixed with equal volume of Freund's complete adjuvant/mouse. The subsequent injections consisted of the mixture 100 μ g of ADESH and equal volume of incomplete Freund's adjuvant/mouse. The mice were injected intramuscularly (IM) six times two weeks apart. At the end of the immunization the mice were bled through the ophthalmic veins and serum was separated.

Enzyme-Linked Immunosorbent Assay (ELISA): The binding affinity of Anti-ADESH made against ADESH consisting of ten amino acids (AD-10), to various specimens known to contain NGF, such as venoms, body fluids, saliva, serum, urine etc. was studied by ELISA. The ELISA binding of Anti-AD-10 was compared to Anti-NGF made against cobra venom. ELISA tests were performed in 96 well microtiter plate. The wells of the plate were coated with one concentration of antigen, diluted in PBS each well receiving 100 μ L. The plate was incubated at room temperature (RT) for 16 to 18 hours after which it was three times (3X) with PBS. The wells of the plate were blocked with 0.25ml/well of 3% Teleostean gelatin from cold water fish (Sigma) for 1/2 hour at RT. Anti-AD-10 and Anti-NGFs were diluted threefold in gelatin were added to the appropriate wells of ELISA plate, including positive and negative controls. The plate was incubated at 37° C for 1 to 1.5 hours. After washing 3X times horseradish peroxidase conjugated with IgG made in goat (Sigma) was added and incubated for 1 hour. Finally, the plate was washed and reacted with O-Phenylenediamine Dihydrochloride (OPD) for color development. The test was after 1/2 hour for ELISA titers.

Isolation of NGF from Body Fluids: Concentrated body fluids such as saliva, serum and urine were fractionated on HPLC by our proprietary procedure. Each type fluid resolved into several fractions. The fractions were dialyzed and tested for neurotrophic activity of PC12 cells. The identified fraction of NGF was further repurified to obtain homogenous NGF. NGFs were also isolated from cobra snake serum and from honey bee venom.

Isolation of NGF from Established Eukaryotic Cells: The cells grown in tissue culture medium DMEM was concentrated before fractionating on HPLC, for isolation of NGF. Each type of cell medium resolved into several fractions. The fractions were dialyzed and tested for neurotrophic activity of PC12 cells. The identified fraction of NGF was further repurified to obtain homogenous NGF. The cell cultures used were Chang liver, Vero, PC12, Neuroblastoma and SP/2 cells and the procedure for isolation followed was similar as described above.

RESULTS

Table I
Biological Properties of Different Versions of ADESH
compared to Venom NGF

Specimen	Neurites on PC12	Toxicity to PC12	Source	#amino acids	Mol. wt.
V- NGF	5 ng	5 μ g	Venom	16	13,500
AD - 15	1 μ g	>100 μ g	Synthetic	15	1,921
AD - 10	1 μ g	>100 μ g	Synthetic	10	1230
AD -5	2 μ g	>100 μ g	Synthetic	5	640

The results of Table 1 show that

- (1) the venom derived NGF is toxic at the concentration of 5 μ g/ml while AD - 15, AD-10 and AD-5 are not toxic up to 100 μ g/ml on PC12 cells.
- (2) Venom derived NGF produces neurite outgrowth at 5 ng/ml on PC12 while each AD-15, AD-10 and AD-5 requires 1000 ng/ml, 200 times the concentration of venom NGF.
- (3) AD-5, AD-10 and AD-15 mimic the property of whole natural NGF in producing neurites on PC12 cells. These properties indicate that AD-5, AD-10 and AD-15 is an integral part of the whole molecule NGF.

Table II
Immunological Properties of AD-15, AD-10 and AD-5:
ELISA titer for Binding Affinity to Anti-AD-10, Anti-V-NGF
and Anti-H-NGF

5	Specimen	Anti-AD-10	Anti-H-NGF	Anti-H-NGF
	V-NGF	900	24300	1800
	AD-15	1800	600	900
	AD-10	2700	450	900
	AD-5	600	300	600
10	H-NGF serum	2700	2700	24300
	H-NGF saliva	2700	1800	8100
	H-NGF urine	2700	1800	8100

Results of Table II show that

- (1) Anti-AD-10 reacts with NGFs derived from venom, human body fluids; serum, saliva and urine, AD-15 and AD-5.
- (2) Binding affinity of Anti-AD-10 is greater to the human source NGFs than venom NGF.
- (3) Anti-V-NGF reacts poorly to AD-15, AD-10 and AD-5 in comparison to Anti-H-NGF.

The binding property of Anti-AD-10 to the natural source NGFs illustrates that AD-10 is an integral part and closer to human NGF.

Table III
ELISA Titers of
Anti-AD-10 and Anti-V-NGF to Venoms

Specimen	Anti-AD-10	Anti-V-NGF
<i>C. atrox</i>	300	900
<i>N. n. kaouthia</i>	600	5400
<i>D. russelli</i>	450	1800
<i>O. Scutellatus</i>	450	1800
Honey Bee	300	2700
Scorpion	300	2700

It is known that snake venoms contain NGF and recently Lipps (1999) has reported that honey bee and scorpion venoms also contain NGF. Results of Table III demonstrate that

(1) Anti-ADESH show binding affinity to venoms similarly to Anti-V-NGF.

(2) This property illustrates that synthetic ADESH consisting of ten amino acids is an integral part of V-NGF which has greater than 60 % homology to human NGF.

Table IV

**ELISA Binding Affinity of Anti-ADESH, Anti-Venom NGF
and Anti-Human NGF to NGFs from Various Sources.**

Cell Type	Origin	Anti-ADESH	Anti-H-NGF	Anti-V-NGF
Chang	human	8100	16200	2700
NB	human	8100	16200	2700
PC12	rat	200	300	2700
Vero	monkey	900	300	2700
SP2	mouse	900	1800	600

Results of Table IV illustrate that:

- (1) Anti-ADESH has highest binding affinity to NGFs derived human source (Chang, NB cells), lesser to monkey and mouse (Vero, SP/2), and least to rat (PC12) cell derived NGF.
- (2) The binding affinity of Anti-ADESH is similar to Anti-H-NGF. This shows that a synthetic peptide ADESH is closer to human NGF is pleasing as ADESH is proposed to treat humans.

---SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANTS: BINIE V. LIPPS AND FREDERICK W. LIPPS
(ii) TITLE OF INVENTION: SYNTHETIC PEPTIDE FOR
NEUROLOGICAL DISORDERS

(iii) NUMBER OF SEQUENCES: 4

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(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: 3.5" FLOPPY DISK, 1.44 MB
(B) COMPUTER: IBM COMPATIBLE
(C) OPERATING SYSTEM: MS DOS 7.1/ WINDOWS 98
(D) SOFTWARE: WORDPERFECT 5.1 FOR WINDOWS

(vi) CURRENT APPLICATION DATA:

(A) APPLICATION NUMBER:
(B) FILING DATE:
(C) CLASSIFICATION: PRELIMINARY CLASS

(vii) ATTORNEY/AGENT INFORMATION:

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(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 116

(B) TYPE: AMINO ACID

(C) STRANDEDNESS: SINGLE

(D) TOPOLOGY: LINEAR

(ii) MOLECULE TYPE: PROTEIN IN SEQ ID NO: 1

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE:

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE: COBRA VENOM

(A) ORGANISM: NAJA NAJA

(B) STRAIN: WILD

(C) INDIVIDUAL ISOLATE:

(D) DEVELOPMENTAL STAGE: ADULT

(E) HAPLOTYPE:

(F) TISSUE TYPE:

(G) CELL TYPE:

(H) CELL LINE:

(I) ORGANELLE:

(vii) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

NH2-

Glu-Asp-His-Pro-Val-His-Asn-Leu-Gly-Glu-His-Pro-Val-Cys-Asx-
Ser-Thr-Ash-Thr-Trp₂₀-Val-Gly-Val-Lys-Thr-Thr-Ala-Thr-Asn-Ile-
Lys-Gly-Ala-Ser-Val-Ser-Val-Met-Glu-Asn₄₀-Val-Asn-Leu-Asp-Asn-
Lys-Val-Tyr-Lys-Gln-Tyr-Phe-Phe-Glu-Thr-Lys-Cys-Arg-Asx-Ser₆₀-

Asx-Pro-Pro-Glx-Pro-Gly-Cys-Lys-Gly-Ile-Asx-Thr-Glx-His-Trp-
 Asx-Ser-Tyr-Cys-Thr₈₀-Thr-Ser-Asn-Ser-Phe-Ile-Lys-Ala-Leu-Thr-
 Met-Asx-Glx-Gly-Gln-Ser-Ala-Trp-Arg-Phe₁₀₀-Ile-Arg-Ile-Gix-Thr-
 Ala-Cys-Val-Cys-Val-Ile-Thr-Lys-Lys-Gly-Asn-
 COOH

(3) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 15

(B) TYPE: AMINO ACID

(C) STRANDEDNESS: SINGLE

(D) TOPOLOGY: LINEAR

(ii) MOLECULE TYPE: PEPTIDE IN SEQ ID NO: 2

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE:

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE: SYNTHETIC

(A) ORGANISM:

(B) STRAIN:

(C) INDIVIDUAL ISOLATE:

(D) DEVELOPMENTAL STAGE:

(E) HAPLOTYPE:

(F) TISSUE TYPE:

(G) CELL TYPE:

(H) CELL LINE:

(I) ORGANELLE:

(vii) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

N L G E H P V C D S T D T W V

(4) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 10

(B) TYPE: AMINO ACID

(C) STRANDEDNESS: SINGLE

(D) TOPOLOGY: LINEAR

(ii) MOLECULE TYPE: PEPTIDE IN SEQ ID NO: 3

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE:

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE: SYNTHETIC

(A) ORGANISM:

(B) STRAIN:

(C) INDIVIDUAL ISOLATE:

(D) DEVELOPMENTAL STAGE:

(E) HAPLOTYPE:

(F) TISSUE TYPE:

(G) CELL TYPE:

(H) CELL LINE:

(I) ORGANELLE:

(vii) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

N L G E H P V C D S

(5) INFORMATION FOR SEQ ID NO: 4:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 5

(B) TYPE: AMINO ACID

(C) STRANDEDNESS: SINGLE

(D) TOPOLOGY: LINEAR

(ii) MOLECULE TYPE: PEPTIDE IN SEQ ID NO: 4

(iii) HYPOTHETICAL: NO

(iv) ANTI-SENSE:

(v) FRAGMENT TYPE:

(vi) ORIGINAL SOURCE: SYNTHETIC

(A) ORGANISM:

(B) STRAIN:

(C) INDIVIDUAL ISOLATE:

(D) DEVELOPMENTAL STAGE:

(E) HAPLOTYPE:

(F) TISSUE TYPE:

(G) CELL TYPE:

(H) CELL LINE:

(I) ORGANELLE:

(vii) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

N L G E H

CLAIMS

What is claimed is:

1. A composition of matter comprising ADESH is a synthetic peptide consisting of ten amino acids having sequence of N L G E H P V C D S mimics the biological properties of the whole NGF molecule derived from venom.
2. The synthetic peptide ADESH mimics the biological properties of the intact natural NGF, particularly in regards the stimulation of neurite outgrowths on PC12 cells.
3. The binding affinity of Anti-ADESH to NGFs from human body fluids and human origin eukaryotic cells is higher than Anti-V-NGF, which illustrates that the composition of ADESH consisting of ten amino acids is a conserved domain of the activity of human NGF. Therefore, ADESH is immunologically closer to human NGF than venom V-NGF.
4. Synthetic ADESH has potential for treatment of neurological disorders and can be given by various routes; including injection and orally to reach the brain as a small molecule without blood-brain barrier problem.
5. Because ADESH is immunologically closer to human NGF, Anti-ADESH has potential to assay NGF levels in body fluids such as saliva and urine for diagnostic purposes without the necessity of extracting blood.
6. A composition of matter comprising a peptide consisting of at least the first five amino acids from the N-terminal of the sequence N L G E H P V C D S T D T W V and no more than 25 amino acids.

7. A composition of matter as in claim 6 wherein the peptide containing 5 to 20 amino acids which mimics the biological properties of the intact NGF, particularly in regards the stimulation of neurite outgrowth.
8. Anti-ADESH showed more binding affinity to NGFs from human body fluids and human eukaryotic cells, showing closeness to human NGF.
9. A method of therapy wherein a patient is a victim of a neurogenerative disease, particularly Alzheimers or Parkinson's disease; and ADESH is administered by any of various routes including: nasal insufflation, buccal cavity administration, oral ingestion, intramuscular or intravenous injections.
10. A composition of matter comprising of antibody made versus ADESH is claimed for diagnostic use for assaying NGF levels for various phases of neurological disorders and microgravity environment.
11. A composition of matter comprising of antibody made versus peptides containing from 5 to 15 amino acids from the N-terminal of the sequence N L G E H P V C D S T D T W V for assaying NGF levels.
12. A composition of matter comprising a peptide consisting of at least the first five amino acids from the N-terminal of the sequence N L G E H P V C D S T D T W V and no more than 25 amino acids total.
13. A composition of matter as in claim 12 wherein the peptide contains no more than 15 amino acids total.
14. A synthetic peptide which produces an antibody which has a binding affinity to NGFs from human body fluids and human origin eukaryotic cells which is higher than a binding

affinity exhibited by an antibody produced in immunological response to an NGF derived from venom.

15. A method for administering a nerve growth factor to a patient in need of such treatment, said method comprising

selecting a nerve growth factor having in the range of 5 to 20 amino acids, and capable of crossing the blood-brain barrier, and

administering said nerve growth factor to said patient in a manner to reach the bloodstream of the patient.

16. A method as in claim 15 wherein the patient is a victim of a neurodegenerative disease selected from the group consisting of Alzheimer's disease and Parkinson's disease and the administration technique is selected from the group consisting of nasal insufflation, buccal administration, oral ingestion, and intramuscular injection.

17. A method as in claim 16 wherein the nerve growth factor comprises a peptide consisting of

at least the first five amino acids from the N-terminal of the sequence N L G E H P V C D S T D T W V and no more than 25 amino acids total.

18. A process comprising contacting, in vitro, a human nerve growth factor with an antibody made against a peptide containing at least five amino acids from the N-terminal of the sequence N L G E H P V C D S T D T W V and no more than 25 amino acids total.

19. A process as in claim 18 wherein the contacting is carried out so as to cause the antibody to react immunologically with the human nerve growth factor.

ABSTRACT OF THE DISCLOSURE

The purified nerve growth factor consisting of 116 amino acids from the venom of Naja kaouthia snake was fragmented by trypsin digestion. The fragments were isolated individually by high pressure liquid chromatography (HPLC). Thus separated fragments were tested for the biological activity of neurite growth on rat adrenal pheochromocytoma (PC12) cells. The fragment which showed the most activity was named ADESH. Subsequently, ADESH was sequenced. Synthetic ADESH was constructed using ten amino acids N L G E H P V C D S of the fragment from its N-terminal is designated as AD-10. Different versions of synthetic ADESH such as AD-15 and AD-5 consisting of 15 and 5 amino acids respectively were constructed; having the sequence: N L G E H P V C D S T D T W V for AD-15 and N L G E H for AD-5. The synthetic AD-15 and AD-5 mimic the biological activity of the natural NGF.

Synthetic ADESH is equally active as the fragment of the native NGF. Antibodies versus ADESH react with natural NGF having 116 amino acids. However, Anti-NGF reacts poorly with ADESH. This illustrates that the ten amino acids of ADESH are essentially important for the biological activity of neurite growth. Therefore, synthetic ADESH consisting of ten amino acids is a candidate for the treatment of neurological disorders instead of the entire NGF molecule. Synthetic ADESH can be immensely useful for treating neurological disorders such as Alzheimer's disease (AD) and Parkinson's disease (PD). ADESH being a small molecule will overcome the hindrance of blood-brain barrier and can be delivered by injection, as well as by nasal insufflation, buccal cavity, etc. It can be produced cheaply and abundantly with batch to batch reproducibility.

COMBINED DECLARATION AND POWER OF ATTORNEY
(JOINT INVENTORS)

As a below named inventor, I hereby declare that:

TYPE OF DECLARATION

This declaration is of the following type:

☒ original

INVENTOR IDENTIFICATION

My residence, post office address and citizenship are as stated below next to my name. I believe I am an original, first and joint inventor of the subject matter which is claimed and for which a patent is sought on the invention entitled:

TITLE OF INVENTION

SYNTHETIC PEPTIDE FOR NEUROLOGICAL DISORDERS

SPECIFICATION IDENTIFICATION

the specification for which is attached hereto.

ACKNOWLEDGMENT OF REVIEW OF PAPERS AND DUTY OF CANDOR

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information

- which is material to patentability as defined in 37, Code of Federal Regulations, § 1.56.

POWER OF ATTORNEY

I hereby appoint the following attorney(s) and/or agent(s) to prosecute this application and transact all business in the Patent and Trademark Office connected therewith:

John R. Casperson, Reg. No. 28,198.

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DECLARATION

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code, and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

SIGNATURE(S)

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Given Name

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Last Name

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